

the pain or discomfort. Detection of cows in that stage is possible using newer technologies such as infrared thermography. Automated lameness detection systems are very useful, as farmers do not have to spend time watching cows walk. For that purpose can be used a pressure platforms that can measure the cows pressure (separately for each hoof) when the cow walks or stands (Pastell *et al.*, 2010; Maertens *et al.*, 2011). In addition, there are systems that allow automatic tracking the changes in feeding behavior and activity associated with lameness (Blackie *et al.*, 2011; Beer *et al.*, 2016). According to previous research, it was observed the necessity of using multiple monitoring systems, i.e. different sensors, in order to increase accuracy of problem detection (Borchers *et al.*, 2017). Many automated lameness detection technologies have been proposed to assist dairy farmers in managing their herds. However, reasons such as cost and skepticism among farmers themselves, who do not see the usefulness of these new technologies for application on their farms, are limiting factors for the greater application of such innovative systems on farms (Dutton-Regester *et al.*, 2020). Dutton-Regester *et al.* (2020) also points out that is a need to more thoroughly evaluate the effectiveness of these technologies under on-farm conditions in order to generate the necessary data required to show dairy farmers that these technologies are reliable and are economically rational for their dairy business.

ADVANTAGES OF INFRARED THERMOGRAPHY

Infrared thermography (IRT) is a contactless method that measures emitted infrared radiation of the surface temperature of an object (Alsaod *et al.*, 2015). Lin *et al.* (2018) states that IRT is such tip diagnostic tool which can detect minor changes in skin temperature without touching the animals. Which can indirectly measures blood flow changes associated with inflammation from foot lesions. This method has been widely used in industry, veterinary medicine and in livestock production (Hurnik *et al.*, 1985; Harper, 2000; Eddy *et al.*, 2001). Furthermore, according to Alsaod and Büscher (2012) and Bobić *et al.* (2017) this method could be used as tool for the prevention of lameness in dairy production. That support also Schaefer and Cook (2013), who emphasizes that there are compelling thermal signatures that can be identified in the image when lameness occurs due to inflammation. It is known that during inflammation, vascular circulation increases, which consequently leads to increased blood flow to the inflamed area, thereby changing the temperature of the inflamed part of the body. The temperature of extremities and skin is largely dependent on the underlying circulation and tissue metabolism rate. Inflammation or metabolic activity can be associated with changes in blood flow which lead to changes in the amount of heat radiated and consequently with surface thermal patterns, which can be easily identified using infrared thermography (Bobić *et al.*, 2024). Nikkhah *et al.* (2005) concluded that IRT could detect inflammation due to laminitis in cows during early and mid lactation. In the research of the Alsaod *et al.* (2014) was conducted a threshold value of 0.99 °C in maximum coronary band temperature difference between hind and front feet for detection of dermatitis digitalis. According to Harris-Bridge *et al.* (2018) the maximum temperature measured at the heels had the highest accuracy in detecting and predicting

of lameness despite of the risk of contamination through dirt and faeces in housed dairy cattle. Furthermore, the opposite opinion was held by Marti *et al.* (2015) who consider that infrared thermography was not a useful tool for differentiating of the claw area temperatures between beef feedlot cattle diagnosed with foot rot and digital dermatitis. Another study that also gave a positive opinion about thermal imaging is a study from LokeshBabu *et al.* (2018) which emphasises that the infrared thermography is a method with high rate of success methods for early detection of lesions before the occurrence of visibly symptoms. And they also add that infrared thermography have good results especially in combination with other diagnostic tools.

DISADVANTAGES OF INFRARED THERMOGRAPHY

Each diagnostic method has its advantages but unfortunately also certain disadvantages. One of the main disadvantages of infrared thermography is the susceptibility to environmental conditions. The accuracy and the interpretation of thermograms of thermal imaging is determined by many things, such as: ambient temperature, current environmental conditions, solar radiation, airflow, the size of the scanned object and background radiation, angle and distance (Alsaad and Büscher, 2012; Westermann *et al.*, 2013; McManus *et al.*, 2022). Besides that, the influence of the animal itself is great, because of the different blood circulation and body status of ich animal can influence the thermal radiation thus complicating the interpretation of thermograms (Alsaad *et al.*, 2015). Body temperature varies throughout the day and depends on different activities of the animal (physical activity, food intake, time of day etc. (Novotna *et al.*, 2019; Moreira *et al.*, 2021). In addition to the influence of animals, environmental factors also affect the appearance of the thermogram, for exemple by dirt that covers the animal, which impairs the normal emissions of infrared radiation from the body surface. Montanholi *et al.* (2015) state that image quality can be affected by the distance from which it is captured. The choice and quality of thermocamera is very important, because the infrared thermography sometimes gives false positive results and can lead to wrong conclusions (Novotna *et al.*, 2019). High-quality thermocameras are expensive, but cheaper models are available on market have lower image quality and resolution (Lokesh-Babu *et al.*, 2018). Therefore, it is necessary to select suitable equipment for thermographic measurements (Montanholi *et al.*, 2015; Werema *et al.*, 2023), and take into account all the possible negative influences that can lead to an incorrect reading of the thermogram.

CONCLUSIONS

Infrared thermography has great potential as an automated diagnostic method for lameness in dairy cows. Due to the great influence of the environment and the animal itself, it is crucial to ensure adequate conditions at the location of the recording and to consider the influence of the animal itself and its current biological status. It is necessary to have additional support from other lameness detection techniques and data sets that will complement the application of IRT.

REFERENCES

- Alsaad, M., Syring, C., Dietrich, J., Doherr, M. G., Gujan, T., & Steiner, A. (2014). A field trial of infrared thermography as a non-invasive diagnostic tool for early detection of digital dermatitis in dairy cows. *Veterinary journal (London, England : 1997)*, 199(2), 281–285. <https://doi.org/10.1016/j.tvjl.2013.11.028>
- Alsaad, M., Schaefer, A. L., Büscher, W., & Steiner, A. (2015). The Role of Infrared Thermography as a Non-Invasive Tool for the Detection of Lameness in Cattle. *Sensors (Basel, Switzerland)*, 15(6), 14513–14525. <https://doi.org/10.3390/s150614513>
- Alsaad, M., & Büscher, W. (2012). Detection of hoof lesions using digital infrared thermography in dairy cows. *Journal of dairy science*, 95(2), 735–742. <https://doi.org/10.3168/jds.2011-4762>
- Beer, G., Alsaad, M., Starke, A., Schuepbach-Regula, G., Müller, H., Kohler, P., & Steiner, A. (2016). Use of Extended Characteristics of Locomotion and Feeding Behavior for Automated Identification of Lameness in Dairy Cows. *PLoS one*, 11(5), e0155796. <https://doi.org/10.1371/journal.pone.0155796>
- Blackie, N., & Maclaurin, L. (2019). Influence of Lameness on the Lying Behaviour of Zero-Grazed Lactating Jersey Dairy Cattle Housed in Straw Yards. *Animals : an open access journal from MDPI*, 9(10), 829. <https://doi.org/10.3390/ani9100829>
- Bobić, T., Mijić, P., Gregić, M., Baban, M., & Gantner, V. (2016). Primjena termovizijske kamere u ranom otkrivanju bolesti papaka mliječnih krava. *Krmiva*, 58 (2), 55-59.
- Bobic, T., Mijic, P., Gregic, M., & Gantner, V. (2024). Evaluation of the hoof's temperature variations depending on lesion presence, measurement points and leg position. *Veterinarni medicina*, 69(6), 185–190. <https://doi.org/10.17221/8/2024-VETMED>
- Borchers, M. R., Chang, Y. M., Proudfoot, K. L., Wadsworth, B. A., Stone, A. E., & Bewley, J. M. (2017). Machine-learning-based calving prediction from activity, lying, and ruminating behaviors in dairy cattle. *Journal of dairy science*, 100(7), 5664–5674. <https://doi.org/10.3168/jds.2016-11526>
- Cha, E., Hertl, J. A., Bar, D., & Gröhn, Y. T. (2010). The cost of different types of lameness in dairy cows calculated by dynamic programming. *Preventive veterinary medicine*, 97(1), 1–8. <https://doi.org/10.1016/j.prevetmed.2010.07.011>
- Dutton-Regester, K. J., Barnes, T. S., Wright, J. D., & Rabiee, A. R. (2020). Lameness in dairy cows: farmer perceptions and automated detection technology. *Journal of Dairy Research*, 87(S1), 67–71. <https://doi.org/10.1017/s0022029920000497>
- Eddy, A. L., Van Hoogmoed, L. M., & Snyder, J. R. (2001). The role of thermography in the management of equine lameness. *Veterinary journal (London, England : 1997)*, 162(3), 172–181. <https://doi.org/10.1053/tvjl.2001.0618>
- Harris-Bridge, G., Young, L., Handel, I., Farish, M., Mason, C., Mitchell, M. A., & Haskell, M. J. (2018). The use of infrared thermography for detecting digital

- dermatitis in dairy cattle: What is the best measure of temperature and foot location to use?. *Veterinary journal* (London, England : 1997), 237, 26–33. <https://doi.org/10.1016/j.tvjl.2018.05.008>
- Harper, D. L. (2000). The value of infrared thermography in a diagnosis and prognosis of injuries in animals. *Proceedings, Inframation 2000, Orlando, USA*, 115–122.
- Hurnik, J. F., Webster, A. B., & DeBoer, S. (1985). An investigation of skin temperature differentials in relation to estrus in dairy cattle using a thermal infrared scanning technique. *Journal of animal science*, 61(5), 1095–1102. <https://doi.org/10.2527/jas1985.6151095x>
- Green, L. E., Hedges, V. J., Schukken, Y. H., Blowey, R. W., & Packington, A. J. (2002). The impact of clinical lameness on the milk yield of dairy cows. *Journal of dairy science*, 85(9), 2250–2256. [https://doi.org/10.3168/jds.S0022-0302\(02\)74304-X](https://doi.org/10.3168/jds.S0022-0302(02)74304-X)
- Griffiths, B. E., Grove White, D., & Oikonomou, G. (2018). A Cross-Sectional Study Into the Prevalence of Dairy Cattle Lameness and Associated Herd-Level Risk Factors in England and Wales. *Frontiers in veterinary science*, 5, 65. <https://doi.org/10.3389/fvets.2018.00065>
- Lin, Y. C., Mullan, S., & Main, D. C. J. (2018). Optimising lameness detection in dairy cattle by using handheld infrared thermometers. *Veterinary medicine and science*, 4(3), 218–226. Advance online publication. <https://doi.org/10.1002/vms3.104>
- LokeshBabu, D.S., Jeyakumar, S., Vasant, P. J., Sathiyabarathi, M., Manimaran, A., Kumaresan, A., Pushpadass, H. A., Sivaram, M., Ramesha, K. P., Katakaltware, M. A., & Siddaramanna (2018). Monitoring foot surface temperature using infrared thermal imaging for assessment of hoof health status in cattle: A review. *Journal of thermal biology*, 78, 10–21. <https://doi.org/10.1016/j.jtherbio.2018.08.021>
- Marti, S., Wilde, R.E., Moya, D., Janzen, E.D., Jelinski, M.J., Dorin, C.L., et al. (2015). Infrared thermography as a tool to diagnose foot rot and digital dermatitis in feedlot cattle. *Conference Paper*, July 2015.
- Maertens, W., Vangeyte, J., Baert, J., Jantuan, A., Mertens, K.C., Campeneere, S.D., Pluk, A., Opsomer, G., Weyenberg, S.V., & Nuffel, A.V. (2011). Development of a real time cow gait tracking and analysing tool to assess lameness using a pressure sensitive walkway: the GAITWISE system. *Biosystems Engineering*, 110, 29-39.
- McManus, R., Boden, L. A., Weir, W., Viora, L., Barker, R., Kim, Y., McBride, P., & Yang, S. (2022). Thermography for disease detection in livestock: A scoping review. *Frontiers in veterinary science*, 9, 965622. <https://doi.org/10.3389/fvets.2022.965622>
- Montanholi, Y. R., Lim, M., Macdonald, A., Smith, B. A., Goldhawk, C., Schwartzkopf-Genswein, K., & Miller, S. P. (2015). Technological, environmental and biological factors: referent variance values for infrared imaging of the bovine. *Journal of animal science and biotechnology*, 6(1), 27. <https://doi.org/10.1186/s40104-015-0027-y>

- Moreira, M. O., Qu, Y. F., & Wiens, J. J. (2021). Large-scale evolution of body temperatures in land vertebrates. *Evolution letters*, 5(5), 484–494. <https://doi.org/10.1002/evl3.249>
- Novotna, I., Langova, L., & Havlicek, Z. (2019). Risk factors and detection of lameness using infrared thermography in dairy cows – A review. *Annals of Animal Science*, 19(3), 563-578. <https://doi.org/10.2478/aoas-2019-0008>
- Ózsvári, L. (2017). Economic cost of lameness in dairy cattle herds. *J. Dairy Vet. Anim. Res*, 6(2), 283-289. <http://dx.doi.org/10.15406/jdvar.2017.06.00176>
- Pastell, M., Hänninen, L., de Passillé, A. M., & Rushen, J. (2010). Measures of weight distribution of dairy cows to detect lameness and the presence of hoof lesions. *Journal of Dairy Science*, 93(3), 954–960. <https://doi.org/10.3168/jds.2009-2385>
- Sjöström, K., Fall, N., Blanco-Penedo, I., Duval, J.E., Krieger, M. & Emanuelson, U. (2018). Lameness prevalence and risk factors in organic dairy herds in four European countries. *Livest. Sci.* 208, 44–50. <https://doi.org/10.1016/j.livsci.2017.12.009>
- Werema, C. W., Laven, L., Mueller, K., & Laven, R. (2021). Evaluating Alternatives to Locomotion Scoring for Lameness Detection in Pasture-Based Dairy Cows in New Zealand: Infra-Red Thermography. *Animals : an open access journal from MDPI*, 11(12), 3473. <https://doi.org/10.3390/ani11123473>
- Westermann, S., Buchner, H. H., Schramel, J. P., Tichy, A., & Stanek, C. (2013). Effects of infrared camera angle and distance on measurement and reproducibility of thermographically determined temperatures of the distolateral aspects of the forelimbs in horses. *Journal of the American Veterinary Medical Association*, 242(3), 388–395. <https://doi.org/10.2460/javma.242.3.388>
- Whay, H. R., Main, D. C., Green, L. E., & Webster, A. J. (2003). Assessment of the welfare of dairy cattle using animal-based measurements: direct observations and investigation of farm records. *The Veterinary record*, 153(7), 197–202. <https://doi.org/10.1136/vr.153.7.197>

OTKRIVANJE ŠEPAVOSTI KOD MUZNIH KRAVA PRIMJENOM INFRACRVENE TERMOGRAFIJE

Sažetak

Šepavost mliječnih krava jedan je od najvećih problema na farmama za proizvodnju mlijeka. Šepavost uz bol i nelagodu životinja narušava dobrobit. Osim toga, zahtijeva medicinsku skrb i dodatne troškove koji rezultiraju velikim ekonomskim gubicima. Rano otkrivanje upale papaka kod životinja važno je za sprječavanje šepavosti. Infracrvena termografija (IRT) je beskontaktna metoda za bilježenje temperature na površini tijela životinja ili drugih predmeta. IRT ima veliki potencijal kao automatizirana dijagnostička metoda za šepavost kod mliječnih krava. Svaka metoda ima svoje prednosti i nedostatke, pa tako i IRT, pa prije donošenja konačnog zaključka treba uzeti u obzir sve pozitivne i negativne utjecaje. Zbog velikog utjecaja okoliša i trenutnog biološkog statusa životinje, ključno je osigurati odgovarajuće uvjete na mjestu snimanja te uzeti u obzir utjecaj same životinje. Neophodno je imati dodatnu podršku drugih tehnika za otkrivanje šepavosti i skupova podataka koji će nadopuniti primjenu IRT-a, kao što su vizualno promatranje papaka i metode promatranja ponašanja mliječnih krava.

Ključne riječi: šepavost, otkrivanje, infracrvena termografija, mliječne krave, čimbenici rizika